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ENANTIOMERIC SYNTHESIS OF POLYHYDROXYLATED INDOLIZIDINE ANALOGUES RELATED TO CASTANOSPERMINE: 1-DEOXY-7-EPICASTANOSPERMINE AND 1,7-DIDEOXY-7-FLUOROCASTANOSPERMINE¹

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(Received in UK 20 July 1992)

Abstract: (6S,7S,8R,8aR)-6,7,8-Trihydroxyindolizidine (1) and its (7R)-deoxyfluoro analogue (2) was synthesised from 5,8-benzyloxycarbonylimino-5,6,7,8-tetradeoxy-1,2-O-isopropylidene- α -D-gluco-octose (3). The kcy step is an oxidation-reduction sequence. Introduction of a fluoro substituent at C-3 (6) was readily effected by displacement of the 3-trifluoromethanesulfonyl (triflate) group with tris-(dimethylamino)-sulfonium-difluorotrimethylsilicate (TASF).

INTRODUCTION

The discovery that certain naturally occurring mono- and bi-cyclic polyhydroxylated alkaloids are specific and potent inhibitors of several glycosidases², including those involved in the processing of glycoproteins³, has generated considerable interest in analogues of 1-deoxynojirimycin and castanospermine as glycosidase inhibitors and potential therapeutic agents for the treatment of AIDS⁴. Fluorinated analogues of these compounds could be important because like fluorosugars, they could show potential or enhanced inhibitory activity⁵ against DNA and RNA synthesis and potential antitumour and antiviral agents^{5c,6} thereby providing the prospect of an alternative chemotherapeutic agent against HIV and related viruses^{4c,7}. Few synthetic methods exists for the selective functionalization and transformation of the individual hydroxyl groups of these alkaloids. As part of a synthetic programme on analogues of 1-deoxy-7-epicastanospermine (1) and 1,7-dideoxy-7-fluorocastanospermine (2).



RESULTS AND DISCUSSION

5,8-Benzyloxycarbonylimino-5,6,7,8-tetradeoxy-1,2-O-isopropylidene- α -D-gluco-octose⁸ (3) was readily converted to the allo-isomer by an oxidation (\rightarrow 4) - reduction (\rightarrow 5) sequence in about 65% yield. Compound (4) was isolated as the hydrate as was evident from the ¹H- and ¹³C-n.m.r.spectra. The ¹H-n.m.r. spectrum indicated the presence of two hydroxyl groups, the proton of one appearing downfield (δ 7.11) indicating that it is probably fairly strongly hydrogen bonded. Its ¹³C-n.m.r. spectrum also did not show a carbonyl C signal at δ ~210

Reaction of (5) with triflic anhydride followed by displacement of the triflate group with TASF⁹ in cold dichloromethane gave the 3-deoxyfluoro derivative (6). The ¹H-n.m.r. spectrum of (6) was not well resolved but it was observed that the H-3 signal has been deshielded (cf. 5). The presence of a fluorine substituent at C-3 (δ_F -129.7) was further confirmed by its ¹⁹F-n.m.r. spectrum while the ¹³C-¹⁹F couplings ($J_{2,F}$ 32 $J_{3,F}$ 186.4, $J_{4,F}$ 10.0 Hz) were similar to those of 3-deoxy-3-fluoro-D-glucofuranoid derivatives^{9,10}. Additional evidence was provided by the mass spectrum of (6) which displayed the ions m/z 365 [M⁺] and 350 [M⁺-15].

Hydrolysis of the 1,2-acetal group of (6) using Amberlite IR-120 (H⁺) resin proceeded slowly to afford the free furanose (7) as an anomeric mixture. Catalytic hydrogenation of the mixture effected hydrogenation of the benzyloxycarbonyl group followed by intramolecular reductive amination of the C-1 aldehyde to yield (2) (78% yield). Evidence of the structure of (2) was provided by its ¹H-, ¹³C- and ¹⁹F-n.m.r. spectra. Its n.m.r. signals and the ¹H-¹H, ¹⁹F-¹H and ¹³C-¹⁹F couplings of the piperidine ring are consistent with those found in 1,3-dideoxy-3-fluoronojirimycin¹. The ¹H-n.m.r. spectrum showed a low field signal (δ 4.23) containing two triplets, each of intensity 0.5 protons. This was assigned to H-7. The separation between these two triplets (52.9 Hz) is characteristic of a geminal $^{19}\text{F}^{-1}\text{H}$ coupling. That the fluorine substituent at this position is equatorial is reflected by the large $J_{6,7}$ and $J_{7,8}$ couplings (9.0 Hz). The ${}^{4}J$ couplings between F-7 and H-5eq and C-8a (5.0 and 8.0 Hz, respectively) accord with the stereospecificity of such couplings^{5a}, giving further confirmation of the equatorial orientation of the fluoro substituent. The presence of a deoxy group at C-5 was reflected by the H-5eq and H-6 signals at δ 3.17 ($J_{5ax,5ca}$ 11.0 Hz) and 3.90 ($J_{5ax,6}$ 10.4 Hz), respectively. The H-5ax signal overlapped with the H-8a signal at δ 2.7. The signals due to the protons of the pyrrolidine ring are much as would be anticipated. The presence of a deoxy group at C-1 was reflected by the signals at δ 1.52 $(J_{1a,1b} \ 12.0, J_{1b,8a} = J_{1b,2a} = J_{1b,2b} = 9.0 \text{ Hz}, \text{H-1b})$ and 2.05. In addition, accurate mass measurement gave a fragment m/z 175.0997 corresponding to C8H14FNO2 [M+].

Synthesis of 1-deoxy-7-epicastanospermine (1) was carried out by a similar procedure as for (2), by first hydrolysing the acetal group followed by catalytic hydrogenation. The ¹H n.m.r. spectrum showed coupling constants $J_{6,7} = J_{7,8} = 2.7$ Hz, indicative of an axial substituent at C-7.

EXPERIMENTAL

Optical rotations were determined at 22-25°C in 1 dm tube with a Perkin Elmer 141 polarimeter. The ¹H-, ¹³C- and ¹⁹F- n.m.r. spectra (internal Me₄Si) were recorded on a Brucker ACS-300 (300 MHz) or AMX-500 (MHz) spectrometers. E.i. mass spectra (70 eV) were determined with a Micromass VG 7035 spectrometer. Melting points were determined using a Büchi 512 melting point apparatus and are uncorrected. Microanalyses were carried out using a Perkin Elmer 2400 Elemental Analyser. Reactions were monitored by t.l.c. on silica gel 60 F_{254} (Merck) with detection by charring with sulphuring acid. Flash column chromatography was performed on Kieselgel 60 (Merck 230-400 mesh) at 5-10 p.s.i.

5,8-Benzyloxycarbonylimino-5,6,7,8-tetradeoxy-1,2-O-isopropylidene-α-D-octofuranosid-3-ulose-hydrate (4). A solution of (3) (140 mg) in N,N-dimethylformamide (6 ml) and dimethylsulfoxide (0.15 ml) was heated with phosphorus pentaoxide (160 mg) at 65-70°C for 2.5 h. T.l.c. (ethyl acetate-hexane, 1:2) revealed the presence of a slower-moving product. The cooled mixture was poured into ice-water and extracted with ether (3 x 5 ml). The ether extract was washed once with water, dried (Na₂SO₄) and concentrated to give, after flash column chromatography (ethyl acetate-hexane, 1:2), crystalline (3) (110 mg, 75%), m.p. 75-76°C (from methanol), $[\alpha]_D$ +84° (c 0.6, chloroform), v 3470, 3300, 1660 cm⁻¹. ¹H-N.M.R. δ_H 7.11 [s, 1 H, 3-(OH)₂]; 5.78 (d, 1 H, J_{1,2} 3.9 Hz, H-1); 5.0-5.2 (m, 2 H, ArCH₂); 4.20 (d, 1 H, H-2); 4.0-4.2 (m, 2 H, H-5, 3-(OH)₂]; 3.70 (d, 1 H, J_{4,5} 10.0 Hz, H-4); 3.4-3.6 (m, 1 H, H-8a,8b); 1.8-2.2 (m, 4 H, H-6a,6b,7a,7b); 1.40, 1.56 [d, 6 H, C(CH₃)₂]. ¹³C-N.M.R. δ_C. 114.3 (CMe₂); 102.8/103.4 (C-1/C-3); 79.5/79.2 (C-2/C-4); 67.7 (C-5); 67.0 (ArCH₂); 47.0 (C-8); 28.6 (C-6); 27.3, 27.1 [C(CH₃)₂], 23.9 (C-7). Anal. calc. for C₁₉H₂₅NO₈ (395.41): C-57.71; H 6.37, N 3.54%; found: C 57.72; H 6.71; N 3.92.

5,8-Benzyloxycarbonylimino-5,6,7,8-tetradeoxy-1,2-O-isopropylidene- α -D-allo-octose (5). A solution of (4) in aqueous ethanol (3:7) was treated with sodium borohydride (12 mg) over 15 min and stirred for 2 h at room temperature when t.l.c. (ether-hexane, 1:1) revealed a slower moving product. Ethanol was removed by distillation and the aqueous solution was extracted with dichloromethane, dried (Na₂SO₄) and concentrated. Flash column chromatography (ether-hexane-methanol, 2:2:1) of the syrupy residue gave (5), (73 mg, 78%), $[\alpha]_D$ +91° (c 0.95, chloroform). ¹H-N.M.R. (CDCl₃) δ_H 5.73 (d, 1 H, $J_{1,2}$ 3.7 Hz, H-1); 5.0-5.3 (m, 2 H, ArCH₂); 4.61 (m, 1 H, H-2); 3.4-4.2 (m, 6 H, H-3,4,5,8a,8b,3-OH); 1.8-5.3 (m, 4 H, H-6a,6b,7a,7b); 1.35, 1.56 [2 s, 6 H, C(CH₃)₂]. ¹³C-N.M.R. δ_H 112.7 (CMe₂); 103.4 (C-1); 79.8/77.6 (C-2/C-4); 74.9 (C-3); 67.1 (ArCH₂); 59.3 (C-5); 47.0 (C-8); 27.1 (C-6); 26.4, 26.6 [C(CH₃)₂]; 23.8 (C-7). Anal. calc. for C₁₉H₂₅NO₆ (363.40): C 62.80, H 6.93, N 3.85%; found: C 62.35, H 6.52, N 3.44.

5,8-Benzyloxycarbonylimino-3,5,6,7,8-tetradeoxy-3-fluoro-1,2-O-isopropylidene- α -D-gluco-octose (6). A solution of (5) (0.31 g) in a mixture of dry dichloromethane (15 ml) and pyridine (0.5 ml), cooled in an ice-salt bath, was treated with triflic anhydride (0.55 ml). The solution was stirred for 20 min when t.l.c. (ether-hexane, 1:1) revealed only one faster-moving product. The solution was washed successively with dil. HCl, water (thrice) and dried (Na₂SO₄). The solution was filtered, cooled in ice-bath and treated with TASF (0.9 g) for 1 h when t.l.c. (ether-hexane, 1:1) showed the presence of a faster -moving product. The reaction mixture was washed with water, the organic extract dried (Na₂SO₄) and concentrated to give, after flash column chromatography (ether-hexane, 1:1), (2) (0.24 g, 78%), [α]_D +48.5° (c 0.9, chloroform). ¹H-N.M.R. (CDCl₃) δ _H 5.95 (d, 1 H, J_{1,2} 3.2 Hz, H-1); 5.0-5.1 (m, 2 H, ArCH₂); 4.2-5.0 (m, H-2,3,4,5); 3.4-3.5 (m, 1 H, H-8); 1.8-2.3 (m, 4 H, H-6a,6b,7a,7b); 1.31, 1.59 [2 s, 6 H, C(CH₃)₂]. ¹³C-N.M.R. δ _C 112.2 (s, CMe₂), 104.7 (s, C-1); 94.8 (d, J_{3,F} 186.4 Hz, C-3); 82.6 (d, J_{2,F} 32.0 Hz, C-2); 80.4 (d, J_{4,F} 10.0 Hz, C-4); 66.9 (s, ArCH₂); 55.75 (s, C-5); 46.6 (s, C-8); 26.3, 26.7 [2 s, C(CH₃)₂]; 24.6/23.75 (C-6/C-7). ¹⁹F-N.M.R. δ _F -129.7. Anal. calc. for C₁₉H₂₄FNO₅: C 62.45, H 6.62, F 5.20, N 3.83%; found: C 62.64, H 6.34, F 5.26, N 3.67.

1-Deoxy-7-epicastanospermine (1). A solution of (5) (0.28 g) in aqueous solution (50 ml, 1:10) was stirred with Amberlite IR-120 (H⁺) resin at 45° for 3 h. The solution was concentrated, dissolved in methanol and hydrogenated in the presence of 10% Pd on activated charcoal for 20 hr at room temperature and 35 p.s.i pressure. The catalyst was filtered off and the filtrate concentrated to give, after flash-column chromatography, crystalline (1) (85 mg, 30%), m.p. 123-124° (from methanol-ether), $[\alpha]_D$ +18°+ (c 0.8, methanol). ¹H-N.M.R. $(D_2O) \delta_H 4.04 (t, 1 H, J_{6,7} = J_{7,8} = 2.7 Hz. H-7); 3.89 (qd, 1 H, J_{5eq,6} 5.5, J_{5ax,6} 11.0 Hz, H-6); 3.55 (dd, 1 H, J_{5eq,6} 5.5, J_{5ax,6} 5.5) (dd, 1 H, J_{5eq,6} 5.5, J_{5ax,6} 5.5) (dd, 1 H, J_{5eq,6} 5.5, J_{5ax,6} 5.5) (dd, 1 H, J_{5eq,6} 5.5) (dd, 1 H, J_{5e$ J_{8 8a} 10.1 Hz, H-8); 3.17 (dddd, 1 H, J_{3a.3b} 10.3, J_{2a.3a} 7.3, J_{2b.3a} 4.4 Hz, H-3a); 3.08 (dd, 1 H, J_{5eq.5ax} 11.0 Hz, H-5eq); 2.5-2.8 (m, 2 H, H-1b, 3b, 5ax); 2.0-2.2 (m, 1 H, H-8a); 1.8-2.0 (m, 2 H, H-2a, 2b); 1.4-1.6 (m, 1 H, H-1b). ¹³C-N.M.R. δ_C 73.8/73.4 (C-7/C-8); 69.7 (C-6); 64.1 (C-8a); 55.1 (C-5); 52.35 (C-3); 29.0 (C-1); 22.85 (C-2). Anal. calc. for C₈H₁₅NO₃ (173.21): C 55.47, H 8.73, N 8.09%; found: C 55.86, H 8.54, N 7.76.

1.7-Dideoxy-7-fluorocastanospermine (2). A solution of (5) (140 mg) in methanol (15 ml) was treated as for (1) to give (2) (70 mg, 95%), m.p. 137-138° (from ether), $[\alpha]_D + 44°$ (c 0.9, methanol). ¹H-N.M.R. (D₂O) $\delta_{\rm H}$ 4.23 (dt, 1 H, $J_{6,7} = J_{7,8} = 9.0$, $J_{7,F}$ 52.9 Hz, H-7); 3.90 (qd, 1 H, $J_{5eq,6}$ 5.5, $J_{5ax,6}$ 10.4, $J_{6,7}$ 9.0, $J_{6,F}$ 14.3 Hz, H-6); 3.50 (ddd, 1 H, J8,8a 9.0, J8,F 12.3 Hz, H-8); 3.17 (ddd, 1 H, J5ea.5ax 11.0, J5ea,F 5.0, H-5eq); 2.98 (dt, 1 H, J_{3a,3b} 9.0, J_{2,3eq} 5.5 Hz, H-3eq); 2.1-2.2 (m, 2 H, H-5ax,8a); 2.0-2.1 (m, 1 H, H-1a); 1.79-1.9 (m, 2 H, H-2a,2b); 1.52 (qd, 1 H, $J_{1a,1b}$ 12.0, $J_{1b,8a} = J_{1b,2a} = J_{1b,2b} = 9.0$ Hz, H-1b). ¹³C-N.M.R. δ_{C} 101.9 (d, $J_{7,F}$ 178.7 Hz, C-7); 75.4 (d, J8 F 17.3 Hz, C-8); 71.2 (d, J6 F 17.8 Hz, C-6); 68.8 (d, J8 F 8.0 Hz, C-8a); 56.3 (d, J_{5,F} 9.2 Hz, H-5); 55.3 (s, C-3); 29.9 (s, C-1); 23.9 (s, C-2). ¹⁹F-N.M.R. δ_F -119.3. Anal. calc. for C₈H₁₄FNO₂ (175.20): C 54.85, H 8.05, F 10.84, N 7.99%; found: C 55.22, H 7.64, F 10.44, N 5.59.

Acknowledgments. We thank the National University of Singapore for financial support, and Miss S. Y. Wong, Ms. L. K. Wong and Mr. B. H. Teo for technical support.

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